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Investigation of Zn, Cu, Cd and Hg Concentrations in the Oyster of Chi-ku, Tai-shi and Tapeng Bay, Southwestern Taiwan

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ABSTRACT

Metal concentrations in oyster, *Crassostrea gigas*, were investigated using 155 samples collected seasonally during 1996-2001 from the Chi-ku, Tai-shi, and Tapeng Bay areas in southwestern Taiwan. Seasonal variations as well as site difference were identified in the metal concentrations. The metal concentrations of oyster in southwestern Taiwan indicated a typical seasonal pattern of “winter-spring maximum” and “summer minimum”. Among the three sites, Zn, Cu and Cd contents were found to be the highest in the March sample from Tai-shi, whereas Hg level was found to be the highest in the January sample from Chi-ku. The overall mean concentrations of Zn, Cu, Cd and Hg sampled from the three sites were 860 ± 375, 267 ± 193, 0.954 ± 0.484 and 0.097 ± 0.056 mg/kg dry wt., respectively, representing the baseline metal concentrations of oyster in Taiwan. In comparison with the 1970’s survey, except the 1.8-fold increase in Cu, the other three elements were within the same ranges. After the transformation of the dry-weight-base data into flesh-weight-base data based on a ratio of 6.8 to1, the mean metal levels, for the most part, closely agreed with many international food standards. However, extraordinarily high level of Cu (1115 mg/kg dry wt.) in oyster were found occasionally in wintertime. Therefore, it is strongly suggested that regulations with respect to food safety standards of metal concentrations need to be established as soon as possible.

Key words: baseline, zinc, copper, cadmium, mercury, summer minimum, cultured oyster, Chi-ku, Tai-shi, Tapeng Bay, Taiwan.

INTRODUCTION

Oyster is a major product of inshore mariculture in Taiwan. With a culture area making up over 85% of the total inshore mariculture field, its production, either in terms of quantity or economic value, ranks number one among all mariculture industries in Taiwan(1-3). Its annual production in 1998 to 2000 was NTD 25 to 34 billion consisting of 10-12% of the total aquaculture production in Taiwan. It is one of the most important fishery industries in Taiwan for coastal fishermen. Four western counties, Chiayi, Tainan, Changhua and Yunlin, make up 95% of the total oyster culture in Taiwan.

However, population and commercial activities have grown drastically in these areas over the last few decades. Untreated wastewaters have been allowed to flow through rivers into traditional oyster culture areas at estuary and into shallow coastal waters, thus often resulting in severe incidents of oyster pollution. The most notable case was the green oyster found at the Erhjen Chi estuary in 1987, which was attributed to acid-washed wastewaters from the nearby metal scraping and recycling factories(4). Since the oyster has a high resistance to pollution, they are able to bioaccumulate various metals to a very high degree(4-5).

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In comparison with other coastal areas of Taiwan, the traditional oyster culture areas of Chi-ku, Tai-shi and Tapeng Bay, in Tainan, Yunlin and Pingtung respectively, are all relatively less industrialized and less urbanized. However, plans have been made, and in some cases projects completed for industrial development and national recreational ocean parks. This study was conducted to establish not only baseline data for the assessment of the environmental impact of these projects but also a database for the estimation and evaluation of acceptable safe metal concentrations in oyster in Taiwan.

MATERIALS AND METHODS

I. *Materials*

(I) Collection of specimens

A total of 155 oyster samples separated into sixteen batches were collected from Chi-ku, Tai-shi and Tapeng Bay along the southwestern coast of Taiwan (Figure 1). The oyster samples were collected from Chi-ku and Tapeng Bay during 1996 to 1997 and 1999 to 2000, respectively, and those from Tai-shi during 1998 to 2001.

Once the soft tissues of the oysters were removed, they were thoroughly washed with double distilled water. At least 30 individual oysters were randomly picked out

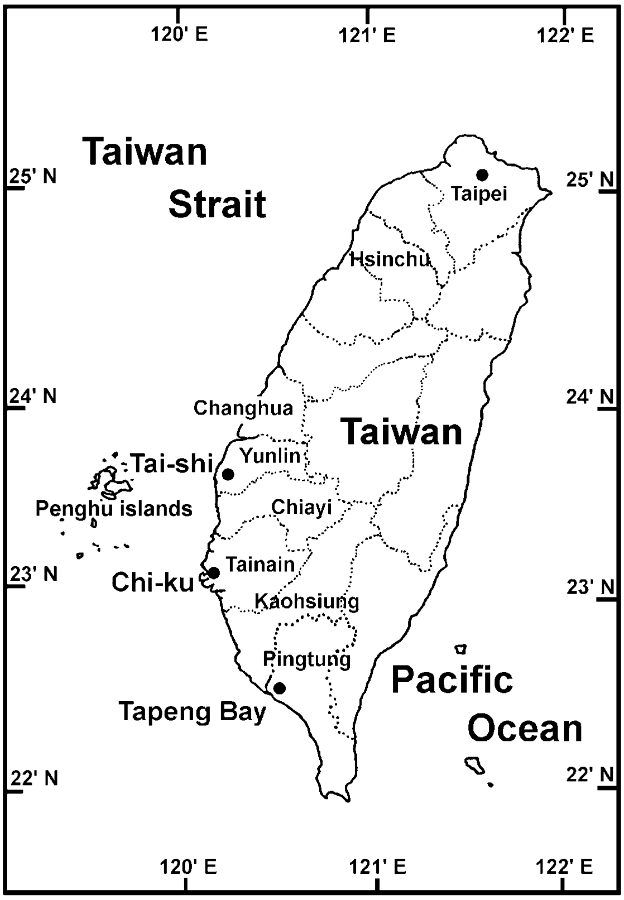

Figure 1. Map of the sampling sites in this study.

from each sample group and combined to make up a pooled sample[6]. Then, the oysters were homogenated and freeze-dried in an acid-washed, clean, white, plastic bottle. The flesh weights and dry weights of the samples were recorded.

(II) *Reagents*

All the chemicals used in this study, including HNO_3, H_2SO_4, $SnCl_2$, $KMnO_3$, were of GR grade, and including the Zn, Cu, Cd and Hg standard solutions of 1000 mg/L, were purchased from the Merck Company. Matrix matched standard reference materials, NIST SRM oyster 1566a from the National Standard Bureau, USA and DORM-2 dogfish muscle from the National Research Council of Canada, were used to control the analytical quality.

II. *Methods*

(I) *Digesting methods*

1. Sample digestion for Cd, Cu and Zn analysis

The sample digestion method for the analysis of Zn, Cu and Cd in this study was the same as in previous studies[7-8], except that the amount of sample and acid were adjusted on the basis of the nature of the sample. In this study, 0.5 g freeze-dried oyster samples were placed into 125 mL of conical flasks, and 15 mL nitric acid was added for digestion. After the digestion, a final volume of 25 mL was made up. Compared with fish tissues, oyster tissues contain a higher percentage of fat requiring longer time to breakdown the organic matter before complete digestion is achieved.

2. Sample digestion for Hg analysis

The methods employed in this study for total mercury analysis were also adopted from previous studies[9-11]. Briefly, 1 g of the freeze-dried oyster tissues was weighed and placed into 75 ml graduated test tubes. 1 mL of HNO_3 and 4 mL of H_2SO_4 were added and heated to 80-90˚C for 1 hour to ensure the tissues completely dissolved. Then 15 mL of 5% $KMnO_4$ was added for the final breakdown of the organic matter in the samples. Finally, the volumes were increased to 25 mL with the addition of double distilled water, and the samples were finally analyzed within 24 hours.

(II) *Analysis of heavy metal concentrations*

1. Measurements of Cd, Cu and Zn

A flame atomic absorption spectrophotometer (Hitachi Zeemen-8200) was used to measure Cu and Zn concentrations in the digested samples. However, Cd was measured using the standard addition method in the atomic absorption spectrophotometer with a graphite furnace. Details of the method are described in Chen and Chen[7].

2. *Measurement of Hg*

The analysis of Hg was carried out using a cold vapor atomic absorption spectrophotometer following the modified system established in Chen's laboratory[9]. 2% of $SnCl_4$ was used as the reductant. The measurement was performed using the cold vapor-AAS method with an Hitachi Z-8200 AAS plus HFS-2 system with a T-joint device.

(III) *Detection limits*

The detection limits were determined following the same method described in previous studies[7,9]. The instrumental detection limits of Zn, Cu, Cd and Hg were 0.02, 0.03, 0.001 and 0.001 mg/L, respectively. The detection limits of Zn, Cu, Cd and Hg in oyster were 1, 1.5, 0.05 and 0.025 mg/kg dry wt., respectively.

III. *Analytical Quality Control and Ensuring Accuracy*

(I) *Quality control measures*

In order to achieve high quality in the analytical results, strict controls were implemented: (1) Each sample

Table 1. Analytical results of standard reference materials (SRM) in this study

Unit: mg/kg

SRM		Zn	Cu	Cd	Hg
Oyster	Certified value	830 ± 57	66.3 ± 4.3	4.15 ± 0.38	0.0642 ± 0.0067
SRM 1566a	This study	858 ± 22	67.2 ± 3.8	4.20 ± 0.10	0.0702 ± 0.0001
	Recovery (%)	103	101	101	109
Dogfish muscle	Certified value	25.6 ± 2.3	2.34 ± 0.16	0.043 ± 0.008	4.64 ± 0.26
DORM-2	This study	23.9 ± 0.9	2.26 ± 0.42	0.046 ± 0.003	—
	Recovery (%)	93	97	107	—

was digested in duplicate in order to determine its heterogeneity. (2) During the experiment, reagent blanks were inserted in every twentieth sample to detect any possible alien containments. (3) Duplicates of the standard reference materials were added simultaneously in each digesting batch. (4) The metal concentrations of NIST oyster 1566a and DORM-2 were then measured to verify the analytical quality.

(II) *Analytical results of the standard reference materials*

In this study, the recovery rate of Zn, Cu, Cd and Hg in the two standard reference materials were all within 100% ± 10% (Table 1).

IV. *Statistical Analysis*

The statistical analysis was performed with the SAS software ANOVA (one-way Analysis of Variance)[12]. The Duncan's multiple range test was also adopted so as to examine the differences in the metal concentrations among species ($p < 0.05$).

RESULTS AND DISCUSSIONS

The results of metal analyses showed that the concentrations of these four elements in the oysters varied seasonally (Figure 2). In Chi-ku oyster sampled in January showed the highest concentrations for all four elements, whereas the November sample contained the lowest Cu and Zn concentrations and the June sample had the lowest Cd and Hg concentrations. In Tai-shi, except for the Zn concentrations in the February samples, all the March and February samples showed either the highest or second highest Cu, Cd and Hg concentrations. In May, at this site, all the elements showed the lowest concentrations. These seasonal fluctuations of metal concentrations in oysters have been well documented[13-16]. The highest Zn and Cu were found at the end of the reproductive cycle of oysters, such as *Crassostrea iridescens*[13-14] and *C. corteziensis*[15] in October in Mexico. During that time, the oyster became skinny due to the repelling of gametes, which meant that Cd and Hg residues in the viscera were concentrated on a mass basis. Similar seasonal variations were also reported by Hsu *et al.*[16] on an oyster study in Yunlin and Chiayi, from southwestern Taiwan in the 1970's. The reproductive cycle of *C. gigas* in southwestern Taiwan varied geographically, demonstrating a trend whereby the oysters at the south of the island commenced their development of gametes earlier than did those of the northern populations, while the October to January period was the resting phase of oysters around the Island[17]. Therefore, the peak metal accumulation season may very well be highly related to the beginning of the reproductive cycle of the oysters. Another factor related to the bioaccumulation of metals in the oyster is the solubility of metal ions in the lower salinity runoff[18], which normally contained more biological sources of metal. In addition, in southwestern Taiwan the dry season normally begins in November and ends in March. During this period of time, except for Zn, concentrations of Fe, Mn, Cu, Ni and Pb in the dissolved phase in water were significantly higher than those during the wet season[19], which enriched the metal concentrations in the coastal waters and possibly caused the metal concentrations to become elevated in March in the Tai-shi samples, as well as in the April samples studied by Hsu *et al.*[16]. In the summer, during the pre-spawning period of oyster, the size and gonad growth in the oyster body diluted the metal concentrations in the whole body[14]. Thus, the lowest metal concentrations were found in May and June.

Table 2 shows the ranges of metal concentrations of oyster analyzed in the current study. For the summer samples, site differences were observed and their trends varied depending on the metal. Among the three sites, samples from Tapeng Bay showed the highest mean Cd and Hg concentrations but the lowest Zn and Cu levels. However, the Chi-ku samples contained the highest Zn and the lowest Cd concentrations, and the Tai-shi samples had the highest mean Cu and the lowest mean Hg (Table 2). Since oysters are known as very sensitive bivalves able to reflect geographical metal gradients in a very short time period[20], the metal concentrations in the summer samples were very low, but they did vary between sites. These fluctuations might be ascribed to natural variations due to geographical differences and environmental conditions.

A further look at the yearly mean concentrations of these metals clearly showed that the differences among the site-year groups were also significant ($p < 0.05$), with the exception of copper (Table 2). This is similar to the summer samples previously discussed, where group differences were metal-dependent. Except a significantly high level of

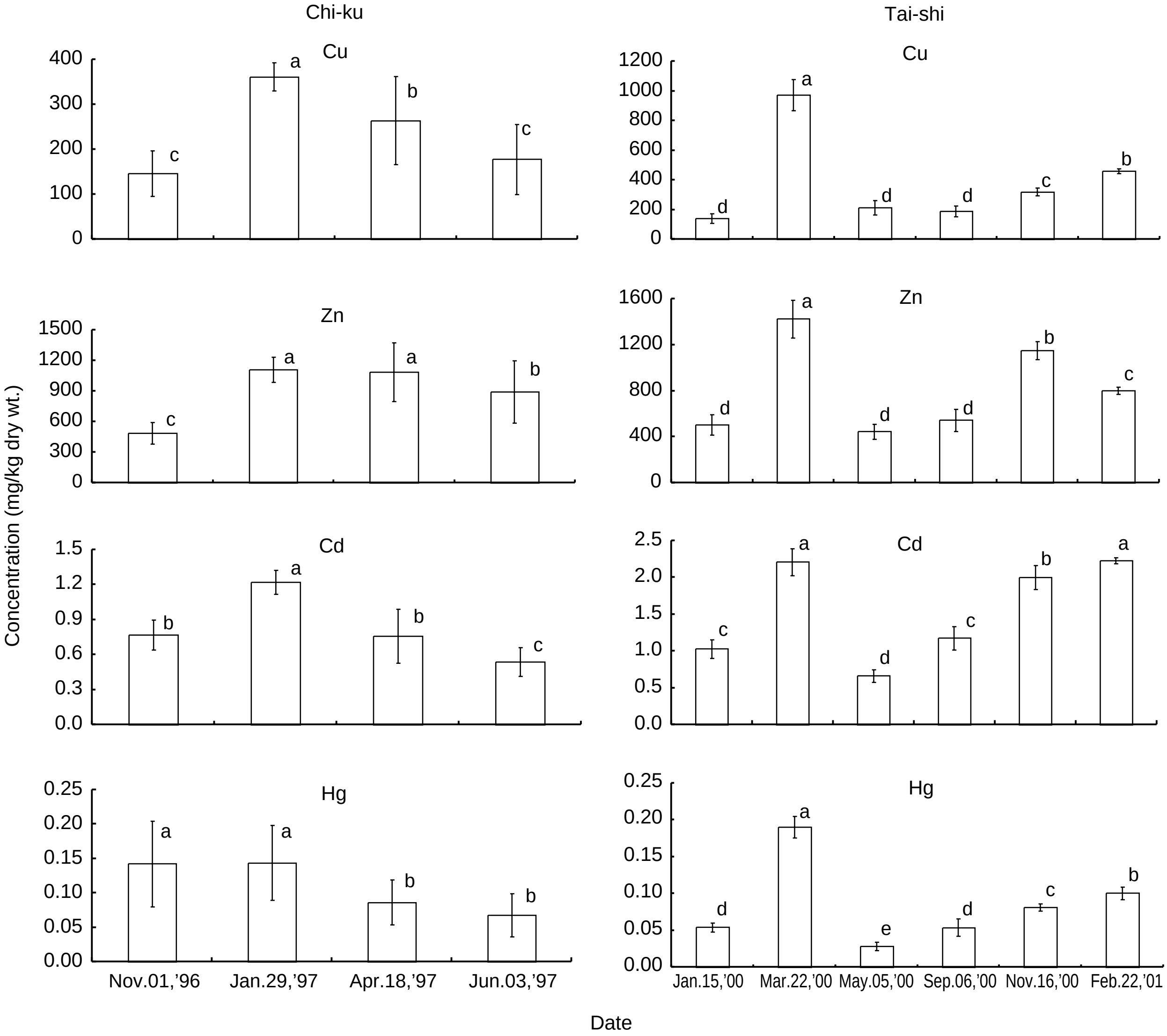

Figure 2. Comparisons of the seasonal variations of metal concentrations of oyster, *C. gigas*, in the Chi-ku and Tai-shi areas, southwestern Taiwan. The bars and vertical lines indicate the means and one standard deviation, respectively. The a, b, c, d and e notations marked above the bars show the results of Duncan's multiple range test at the significant level $p < 0.05$.

Table 2. Mean Zn, Cu, Cd and Hg concentrations (mg/kg dry wt.) in oysters sampled in summer from Tapeng Bay, Chi-ku and Tai-shi and the mean concentrations from whole year samples from Chi-ku and Tai-shi

Site	n	Zn	Cu	Cd	Hg
Summer					
Tapeng Bay	5	260 ±15^{c}	57 ± 17^{b}	0.853 ± 0.130^{a}	0.099 ± 0.028^{a}
		(243~275)	(34~71)	(0.685~0.992)	(0.058~0.132)
Chi-ku	29	893 ± 305^{a}	178 ± 78^{a}	0.540 ± 0.122^{b}	0.068 ± 0.031^{b}
		(437~1281)	(91~321)	(0.312~0.734)	(0.027~0.158)
Tai-shi	15	482 ± 77^{b}	205 ± 46^{a}	0.827 ± 0.245^{a}	0.037 ± 0.015^{c}
		(353~625)	(140~313)	(0.540~1.411)	(< 0.025~0.070)
Whole Year					
Chi-ku (1996-1997)	103	905 ± 332^{ab} (328~1533)	237 ± 108 (72~417)	0.808 ± 0.292^{b} (0.312~1.368)	0.104 ± 0.056^{a} (0.027~0.288)*
Tai-shi (1998-1999)	4	1175 ± 595^{a} (575~1998)	246 ± 53 (184~306)	0.958 ± 0.420^{b} (0.428~1.449)	–
Tai-shi (2000-2001)	34	774 ± 405^{b} (353~1606)	393 ± 319 (114~1115)	1.411 ± 0.683^{a} (0.540~2.452)	0.079 ± 0.059^{b} (< 0.025~0.213)**

Note: The notations a, b, c beside the mean values indicate the results of the Duncan's multiple range test or student's t-test (significant level, $p < 0.05$); and n indicates sample size. Sample size of *: 77 and **: 33.

Hg found in the Chi-ku oysters, the highest mean concentrations of Zn, Cu and Cd were found in the 1998-1999, 2000-2001 and 2000-2001 Tai-shi samples, respectively (Table 2). In comparison with the data reported in the 1970's [16,21], it is interesting to note that a significant doubling of Cu and Cd concentrations were found in the 2000-2001 Tai-shi oyster, which was possibly associated with the extensive construction involved in the Island Industrial Camp off Yunlin. It is well known that digging and dredging of coastal areas greatly contribute to an increase in the precipitation of contaminants in adjacent waters[21], and this could obviously increase the uptake of contaminants by the oyster.

In the current study, the ranking of metal concentrations in oyster was Zn > Cu > Cd > Hg, which reflects the typical metal richness of the *Crassostrea* species, like *C. commercialis*[6], *C. gigas*[5] and *C. corteziensis*[15], as reported in many previous studies[5, 6, 15-16, 21]. The overall mean metal concentrations in the whole soft tissue found in this study (Zn = 860 ± 375, Cu = 267 ± 193, Cd = 0.95 ± 0.48 and Hg = 0.097 ± 0.056 mg/kg dry wt.) were fairly low and comparable to previous records for the coastal waters of Taiwan[4, 5, 16, 21-24], except that the Cu concentration was higher (1.8 folds) than that of 1970's data[16,21] investigating oysters in the same area. In general, the Zn, Cu, Cd and Hg concentrations of oysters in Taiwan were less than 1000, 500, 2.0 and 0.3 mg/kg dry weight. However, for those samples from hot spot areas, such as Hsian-san[5, 23], Charting[4, 23-24] and the Erhjin Chi estuary[4, 23], the Zn and Cu concentrations of oyster reached 2000 and 4000 mg/kg dry weight. It is worth noting, that the Hg concentrations of oysters found in this study were only one-third of the level reported by Jeng *et al.*[23].

Further comparing the metal levels examined in this study with those of the genus *Crassostrea* from other parts of the world found that Zn, Cu, Cd and Hg concentrations in *C. gigas* from southwestern Taiwan are fairly low on a global scale, indicative of typical background level in an agriculture area[25-33]. On a global scale, the Zn, Cu, Cd and Hg concentrations of wild and cultured oysters, were similar to our previous conclusion, less than 1000, 500, 2.0 and 0.3 mg/kg dry weight. A further comparison was made with the background level of wild *Crassostrea* (as *Saccostrea commercialis*) from 27 locations in New South Wales, Australia[33]. It was found that only the highest Cu and Hg concentrations detected in the current study exceeded the 85% of NSWBG (background metal concentrations of oyster in New South Wales, Cu = 170~394 mg/kg dry wt.; Hg = 0~0.1 mg/kg dry wt.) and that the Zn and Cd concentrations were all lower than those in the NSWBG (Zn = 2610~3904 mg/kg dry wt.; Cd = 5~11 mg/kg dry wt.). Our highest Cu level was still within the 85% concentration level of NSWall (all metal concentrations of oyster in New South Wales, Cu = 390~1460 mg/kg dry wt.), and only the highest Hg concentration slightly exceeded the 85% NSWall level (Hg = 0~0.2 mg/kg dry wt.). Accordingly, the metal concentrations of oyster in southwestern Taiwan, as determined in this study, resembled the records of less industrialized and urbanized areas all over the world.

Table 3. International Standards on metal concentrations (mg/kg flesh weight) in seafoods

Country	Standard	Zn	Cu	Cd	Hg	Reference
USA	FDA	–	–	2*	0.5	(34)
USA	NAS	–	–	0.5*	0.5	(35)
Australia	NHMRC	1000	30	2	1.0	(36)
Australia	TPHR	40	30	5.5	1.0	(37)
Canada	-	–	100	-	0.5	(38)
Japan	-	–	–	1	1.0	(38)
UK	MAFF	50	20	-	0.3	(39)
UK	FSC	50	–	-	0.3	(40)

Note: FDA: Food and Drug Administration; NAS: National Academy of Science; NHMRC: National Health Medical Research Council; TPHR: Tasmania Public Health Regulation; MAFF: Ministry of Agriculture, Fisheries and Food; FSC: Food Science Council; and * indicates dry weight base.

After transformation into flesh-weight based on a ratio of 6.8 to 1[34], the highest Cd and Hg concentrations (Cd = 0.361, Hg = 0.031 mg/kg wet wt.) were all within the regulatory food standards of different countries (Table 3). However, the highest Cu level (164 mg/kg wet wt.) was found to exceed all food standards, while the highest Zn (294 mg/kg wet wt.) exceeded most of the standards, with the exception of the NHMRC, Australia[36]. These highest Zn, Cu and Cd concentrations of a single oyster sample were all recorded from the March, 2000 Tai-shi sample, whereas the highest Hg level was from the January, 1997 Chi-ku sample (Table 2). These reflect a geographical and environmental difference within the study area. However, the highest Zn and Cu concentrations were only two-third and one-fourth, respectively, of highest values previously reported[4-5]. The overall mean Zn and Cu levels were 126 ± 55 and 39 ± 28 mg/kg in wet wt., respectively, compliant with the food standards of various countries. So that the metal levels of oysters analyzed in this study represent the metal uptake of the general public through oyster-consumption in Taiwan. Based on these results, it is found that exceptionally high levels of Cu in oyster may occasionally occur in the wintertime. For public health concerns, it is therefore strongly suggested that regulations with respect to food safety standards of metal concentrations, limiting methyl Hg to be below 0.5 mg/kg in wet wt.[41], should also be established for the other elements as soon as possible in Taiwan.

CONCLUSIONS

The investigation into metal concentrations of oyster in southwestern Taiwan showed a typical pattern of "winter-spring maximum" and "summer minimum". The pattern coincides with the reproductive cycle of oyster in the study area, where the spawning of oyster normally occurs in January in the south and July in the north of Taiwan. In addition, the higher levels of metal concentrations in the oyster may be a result of the wet season when larger volumes of freshwater with more bioavailable sources of metal flows down to the estuary. The overall means of Zn, Cu, Cd and Hg concentrations in the oyster were 860 ± 375, 267 ± 193, 0.95 ± 0.484 and 0.097 ± 0.056 mg/kg dry weight, respectively, but the highest Zn, Cu, Cd and Hg concentrations were 1998, 1115, 2.45 and 0.288 mg/kg dry weight, respectively. The metal concentrations represent the metal levels found in a typical agricultural area, the major oyster culture areas and the background levels of oyster in Taiwan. These metal levels in oyster were similar to those reported in the literature. Using a conversion factor of 6.8 from wet to dry wt., high Cu concentrations in oyster were occasionally found in the wintertime, exceeding the regulatory standards of various countries,.

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REFERENCES

1. Anon. 1999. Fisheries yearbook Taiwan area, 1998. Fisheries Administration Council of Agriculture Executive Yuan. p. 378.
2. Anon. 2000. Fisheries yearbook Taiwan area, 1999. Fisheries Administration Council of Agriculture Executive Yuan. p. 400.
3. Anon. 2001. Fisheries yearbook Taiwan area, 2000. Fisheries Administration Council of Agriculture Executive Yuan. p. 419.
4. Han, B. C. and Hung, T. C. 1990. Green oysters caused by copper pollution on the Taiwan Coast. Environ. Pollut. 65: 347-362.
5. Lin, S. and Hsien, I. J. 1999. Occurrence of green oyster and heavy metals contaminant levels in the Sien-San area, Taiwan. Mar. Pollut. Bull. 38: 960-965.
6. Phillips, D. J. H. and Muttarasin, K. 1985. Trace metals in bivalve molluscs from Thailand. Mar. Environ. Res. 15: 215-234.
7. Chen, C. Y. and Chen, M. H. 2001. The heavy metal in nine species of fishes caught in coastal waters off Ann-Ping, S. W. Taiwan. J. Food Drug Anal. 9: 107-114.
8. Smith, S., Chen, M. H., Bailey, R. G. and Williams, W. P. 1996. Concentration and distribution of copper and cadmium in water, sediment, detritus, plants and animals in a hardwater lowland river. Hydrobiologica 341: 71-80.
9. Chen, M.-H. and Chou, C. L. 2000. An instrumental corrective method for the determination of mercury in biological and sediment samples using cold vapor atomic absorption spectrophotometry. J. Chin. Chem. Soc. 45: 1-9.
10. Uthe, J. F., Armstrong, F. A. J. and Stainton, M. P. 1970. Mercury determination in fish samples by wet digestion and flameless atomic absorption spectrophotometry. J. Fish. Res. Bd. Canada 27: 805-811.
11. Uthe, J. F. and Armstrong, F. A. J. 1974. The microdetermination of mercury and organomercury compounds in environmental materials. Toxicol. Environ. Chem. Reviews 2: 45-77.
12. SAS. 1988. Statistical Analysis Software Institute Inc. SAS/STAT®“ User’s, Release 6.03 ed. SAS Institute Inc., p.1028. Cary.
13. Paez-Osuna, F. and Marmolejo-Rivas, C. 1990b. Occurrence and seasonal variations of heavy metals in the oyster, *Saccrostrea iridescens*. Bull. Environ. Contam. Toxciol. 44: 129-134.
14. Paez-Osuna, F., Frias-Espericueta, M. G. and Osuna-Lopez, J. I. 1995. Trace metal concentrations in relation to season and gonadal maturation in the oyster, *Crassostrea iridescens*. Mar. Environ. Res. 40: 19-31.
15. Paez-Osuna, F. and Marmolejo-Rivas, C. 1990a. Trace metal in tropical coastal lagoon bivalves, *Crassostrea corteziensis*. Bull. Environ. Contamin. Toxicol. 45: 538-544.
16. Hsu, S.-Y., Wang, G.-S. and Jeng, S.-S. 1979. The occurrence and seasonal variations of Na, K, Ca, Mg and heavy metals in Taiwan’s oyster and clams. Bull. Inst. Zool., Academia Sinica 18: 11-20.
17. Wu, W. L. and Yang, M. N. 1999. The reproductive cycle of Crassostrea gigas Thunberg, 1793 from Taiwan. Bull. Malacol. Taiwan ROC 23: 39-46.
18. Thomson, J. D. 1982. Metal concentration changes in growing Pacific oysters, *Crassostrea gigas*, cultivated in Tasmania, Australia. Mar. Biol. 67: 135-142.
19. Wann, J. K. and Hung, J. J. 1990. Distribution and transport of heavy metals in the Chishui River. Proc. of the 3rd Conf. on Environmental Planning and Management in the ROC. Nov. 1990. pp. 291-300.
20. Wright, D. A., Mihurshky, J. A. and Phelps, H. L. 1985. Trace metals in Chesapeake Bay oyster: Intra-sample variability and its implication of biomonitoring. Mar. Environ. Res. 16: 181-197.
21. Jeng, S.-S. and Hung, Y.-W. 1973. Heavy metal contents in Taiwan’s cultured fish. Bull. Inst. Zool., Academia Sinica 12: 79-85.
22. Sun, L. T., Huang, S. H. and Chen, H. L. 1986. Heavy metal contents in fish sold from Kaohsiung markets. China Fish. Monthly 403: 9-17.
23. Jeng, M. S., Jeng, W. L., Hung., T. C., Yeh, C. Y., Tseng, R. J., Meng, P. J. and Han, B. C. 1999. Mussel

watch: a review of Cu and other metals in various marine organisms in Taiwan, 1991-1998. Environ. Pollut. 110: 1-9.

24. Lee, C. L., Chen, H. Y. and Chuang, M. Y. 1996. Use of oyster, *Crassostrea gigas*, and ambient water to assess metal pollution status of the Charting coast, Taiwan, after the 1986 green oyster incident. Chemosphere 33: 2505-2532.
25. Silva, C. A. R., Rainbow, P. S., Smith, B. D. and Santos, Z. L. 2001. Biomonitoring of trace metal contamination in the Potengi Estuary, Natal (Brazil), using the oyster, *Crassostrea Rhizophorae*, a local food source. Wat. Res. 35: 4072-4078.
26. Bulter, C. A. and Timperley, M. H. 1996. Fertilized farmland as a source of cadmium in oysters. Sci. Total Environ. 181: 31-44.
27. Hunter, C. L., Stephenson, M. O., Tjeerdema, R. S., Crosby, D .G., Ichikawa, G. S., Goetzl, J. D., Paulson, K. S., Crane, D. B., Martin, M. and Newman, J. W. 1995. Contaminants in oysters in Kaneohe Bay, Hawaii. Mar. Pollut. Bull. 30: 646-654.
28. Pridomore, R. D., Roper, D. S. and Hewitt, J. E. 1990. Variation in composition and condition of the Pacific oyster, *Crassostrea gigas*, along a pollution gradient in Manukau Harbour, New Zealand. Mar. Environ. Res. 30: 163-177.
29. Boyden, C. R. and Romeril, M. G. 1974. A trace metal problem in pond oyster culture. Mar. Pollut. Bull. 5: 74-78.
30. Paez-Osuna, F., Zazueta-Padilla, H. M. and Izaguirre-Fierro, G. 1991. Trace metals in bivalves from Navachiste Lagoon, Mexico. Mar. Pollut. Bull. 22: 305-307.
31. Paez-Osuna, F., Osuna-Lopez, J. I., Izaguirre-Fierro, G. and Zazueta-Padilla, H. M. 1993. Heavy metals in oyster from a subtropical coastal lagoon associated with an agricultural drainage basin. Bull. Environ. Contam. Taoxicol. 50: 696-702.
32. Phillips, D. J. H., Ho, C. T. and Ng, L. H. 1979. The rock oyster *Saccostrea glomerata* as an indicator of trace metals in Hong Kong. Mar. Biol. 53: 353-360.
33. Scanes, P. R. and Roach, A. C. 1999. Determining natural 'background" concentrations of trace metals in oyster from New South Wales, Australia. Environ. Pollut. 105: 437-446.
34. Arnac, M. and Lassus, C. 1985. Heavy metal accumulation (Cd, Cu, Pb and Zn) by smelt (*Osmerus mordax*) from the north shore of the St Lawrence Estuary. Wat. Res. 19: 725- 734.
35. Nabawi, A. E., Heinzow, B. and Kruse, H. 1987. As, Cd, Cu, Pb, Hg and Zn in fish from the Alexandria region, Egypt. Bull. Environ. Contam. Toxicol. 39: 889-897.
36. Bebbington, G. N., Mackay, N. J., Chvojka, R., Williams, R. J., Dunn, A. and Auty, E. H. 1977. Heavy metals, selenium and arsenic in nine species of Australian commercial fish. Aust. J. Mar. Fresh. Res. 28: 277-286.
37. Eustace, I. J. 1974. Zinc, cadmium, copper and manganese in species of fish and shellfish caught in the Denwent Estuary, Tasmania. Aust. J. Mar. Fresh. Res. 25: 209-220.
38. Tsen, J. H. 1996. Investigation on the heavy metal contents of fishes from fish market of Taichung Port. Nutri. Sci. J. 21: 177-188.
39. Sally, E. C., Michael, S. J. and Richard, T. L. 1996. Metal contamination of angler-caught fish from the Mersey Estuary. Mar. Environ. Res. 41: 281-297.
40. Eromosele, C. O., Eromosele, I. C., Muktar, S. L. M. and Birdling, S. A. 1995. Metals in fish from the upper Benue River and Lakes Geriyo and Njuwa in northeastern Nigeria. Bull. Environ. Contam. Toxicol. 54: 8-14.
41. Anon. 1998. Compilation of the laws of food hygiene, February 1998. Sanitation Agency Executive Yuan. p. 378.

5

中國芥茉籽（河北、陝西及山東產地）精油的化學成份

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摘　要

利用氣相層析和氣相層析-質譜聯用技術分析了不同產地（河北、陝西及山東）的中國芥茉揮發油的化學成份。從河北芥茉油中，我們鑒定了22種化合物，佔總精油的89.6%，河北芥茉油的主要成份是：烯丙基異硫氰酸脂（54.8%）、二烯丙基三硫醚（9.4%）、二烯丙基硫醚（5.5%）和3-丁烯基異硫氰酸脂（4.8%）。從陝西芥茉油中，我們鑒定了14種化合物，佔總精油的94.5%，陝西芥茉油的主要成份是：烯丙基異硫氰酸脂（68.8%）、二烯丙基三硫醚（7.8%）和3-丁烯基異硫氰酸脂（4.9%）。從山東芥茉油中，我們鑒定了15種化合物，佔總精油的94.3%，山東芥茉油的主要成份是：烯丙基異硫氰酸脂（61.3%）、二烯丙基三硫醚（9.7%）和3-丁烯基異硫氰酸脂（5.9%）。從所獲得的分析結果來看，中國芥茉油的成份均是由異硫氰酸脂和硫醚所構成的。

關鍵詞：芥茉，烯丙基異硫氰酸脂，芥茉油，精油，氣相層析-質譜儀

7

台灣地區之台西、七股及大鵬灣產牡蠣的鋅、銅、鎘及汞含量之探討

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摘　要

本研究是在1996至2001年間進行依季別自台西、七股及大鵬灣採集當地養殖的牡蠣，總共分析了155個樣品。結果發現牡蠣體內所含之鋅、銅、鎘及汞濃度有地點的差異及季節變化。本研究的牡蠣呈現與前人研究相似的現象，呈現冬季高，夏季低之重金屬濃度的季節變化。三個地點中，最高的鋅、銅及鎘測值出現在台西三月的樣品，而汞則出現在七股元月的樣品。總體而言，三個養殖區的牡蠣所含的鋅、銅、鎘及汞的總平均值及標準偏差分別為860 ± 375、267 ± 193、0.954 ± 0.484及0.097 ± 0.056 mg/kg乾重，與世界其他未受污染地區的牡蠣測值相似，同時，除銅高出1.8倍外，其他三元素亦與1970年代雲嘉養殖區的牡蠣測值相類似。將這些乾重測值經鮮重比乾重為6.8:1換算後，亦皆較世界各國所訂之食品安全限值為低，顯示無食用之安全顧慮。然而，在冬季仍會有偶發性牡蠣銅含量（1115 mg/kg乾重）過高的情形，因此籲請相關食品衛生管理單位應盡速擬訂管理辦法，以保障消費者的健康。

關鍵詞：基礎背景值、鋅、銅、鎘、汞、夏季最低值、養殖牡蠣、七股、台西、大鵬灣、台灣

6

影響*Aspergillus terreus* CCRC 32111於馬鈴薯葡萄糖培養基中產生土震素B之因素

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摘　要

本研究的目的是測定培養時間、溫度、培養基容量及培養基初期酸鹼值對*Apsergillus terreus* CCRC32111產生土震素B之影響。研究結果顯示，以馬鈴薯葡萄糖培養基培養12天及溫度為28°C時，土震素B產量及菌絲生長達最高峰。在黴菌生長停滯期，即培養基中之碳水化合物快速消耗後，土震素B產量達最高量。當培養基容量增加時，土震素B產量下降；但培養基初期酸鹼值為鹼性時，增加培養基容量會使土震素B產量不減反增。由這些結果顯示培養時間、溫度、培養基容量及培養基初期酸鹼值皆會影響土震素B的產生。

關鍵詞：培養時間、溫度、培養基容量、培養基初期酸鹼值、*Aspergillus terreus* CCRC 32111、土震素B、停滯期、碳水化合物

8

原子吸收光譜定量分析安胎劑中鉛、鎘、鉻及砷的含量

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摘　要

本篇使用3種不同的前處理方法與原子吸收光譜來定量安胎劑中所含13種中藥中鉛，鎘，鉻，砷四種元素的含量。乾式灰化，濕式消化，和微波消化3種前處理方法被用來比較安胎劑13種中藥中所含鉛，鎘，鉻，砷之回收率與精密度。乾式灰化，濕式消化，和微波消化的平均回收率分別為80 ± 12%，85.7 ± 3.3%，and 94.5 ± 2.1%而其測量四種元素的平均相對標準偏差分別為8.8 ± 1.7%，7.1 ± 3.3%，and 3.4 ± 0.8%。微波消化前處理與原子吸收光譜對鉛，鎘，鉻，砷的偵測極限分別為0.45 ppb，0.03 ppb，0.20 ppb，and 0.64 ppb而其測量13種中藥中所含鉛，鎘，鉻，砷之平均濃度分別為28.0 ± 0.7 ppb，1.11 ± 0.03 ppb，8.5 ± 0.3 ppb，and 1.52 ± 0.04 ppb。在13種中藥中荊芥含最高之鉛（60.5 ± 2.1 ppb），鉻（23.3 ± 0.5 ppb），及砷（8.8 ± 0.1 ppb）而川芎含最高之鎘（3.76 ± 0.04 ppb）。

關鍵詞：安胎劑，樣品前處理，原子吸收光譜